

Preclinical Studies with HMI-104, an AAVHSC Vectorized C5 Monoclonal Antibody, for the Treatment of Paroxysmal Nocturnal Hemoglobinuria

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Abstract # 386

AAVHSC GTx-mAb Therapy Targeting Complement Mediated Disorders

- Adeno-associated virus (AAV) gene therapy has the potential to offer a long-term resolution for diseases that rely on chronic dosing of therapies such as monoclonal antibody (mAb)-based drugs.
- GTx-mAb platform is an extension of our gene therapy approach that aims to provide systemic and sustained levels of a therapeutic mAb with one-time intravenous (I.V.) dose.
- Previously, we showed that GTx-mAb platform constructs achieved dose-dependent and sustained expression of a functional C5 monoclonal antibody (C5mAb) in NOD SCID mice, and humanized liver FRG® mice,² supporting the use of a vectorized approach for diseases relying on chronic anti-C5 dosing, such as Paroxysmal Nocturnal Hemoglobinuria (PNH).

PNH is a Rare, Acquired, Life-Threatening Blood Disease Treatable with Complement Inhibitors

- PNH is caused by acquired *PIGA* mutations in hematopoietic stem cells, leading to loss of surface binding of the complement inhibitors CD55 and CD59.3
- Blood cells derived from mutant PNH clones expand and are vulnerable to complement-induced hemolysis, leading to major complications such as anemia, thrombosis and bone marrow failure.
- Life expectancy in untreated PNH patients is 10-15 years after diagnosis.^{4, 5}
- Treatment of PNH relies on chronic intravenous (I.V.) dosing of anti-C5 mAbs (eculizumab or ravulizumab every 2 or 8 weeks, respectively), or twice-a-week subcutaneous dosing of an anti-C3 inhibitor (pegcetacoplan) (Figure 1A).

Nonclinical Studies with HMI-104

- HMI-104 is an AAVHSC vector and our developmental GTx-mAb candidate designed to elicit hepatic expression of a C5mAb for the treatment of PNH and alleviate the dependency on chronic dosing of anti-C5 therapeutics (Figure 1A).
- HMI-104 is delivered via a single (I.V.) injection and the expression of C5mAb can inhibit C5mediated lysis of red blood cells (Figure 1B).
- Here, we present the results from two IND-enabling studies with HMI-104:
- -A 16-week dose range finding (DRF) study in NOD SCID mice which lack murine C5 (Figure 2)
- -A 4-week DRF study in humanized liver FRG® KO mice which express human C5 (Figure 3)

Results

16-Week DRF Study of HMI-104 in NOD SCID Male Mice

Objective

To determine the relationship between dose levels, liver vector genomes (vgs), mRNA levels, C5mAb serum concentrations, and ex vivo hemolysis over time.

Results (Figure 2)

- Dose-dependent increase in serum C5mAb levels, with concentrations rising steadily through 5 weeks post dose, and plateauing through 16 weeks post dose
- Linear range for steady state concentrations as a function of dose for Doses B-D Functional C5mAb determined in an ex vivo hemolysis assay using serum from HMI-104treated NOD SCID mice
- Dose-dependent increase in liver vgs, with stable levels achieved by 4 weeks
- Dose-dependent increase in mRNA levels in liver mirroring changes in vgs
- Pharmacologically relevant dose range for HMI-104 in NOD SCID mice established as >A to <E, given that Dose A is considered to be minimally efficacious, and Dose E yielded same C5mAb levels as Dose D

4-Week DRF Study in the Humanized Liver FRG® KO Male Mice

Objective

To evaluate HMI-104 transduction (vgs) and mRNA levels in human hepatocytes in vivo and assess C5mAb levels in the presence of human C5.

Results (Figure 3)

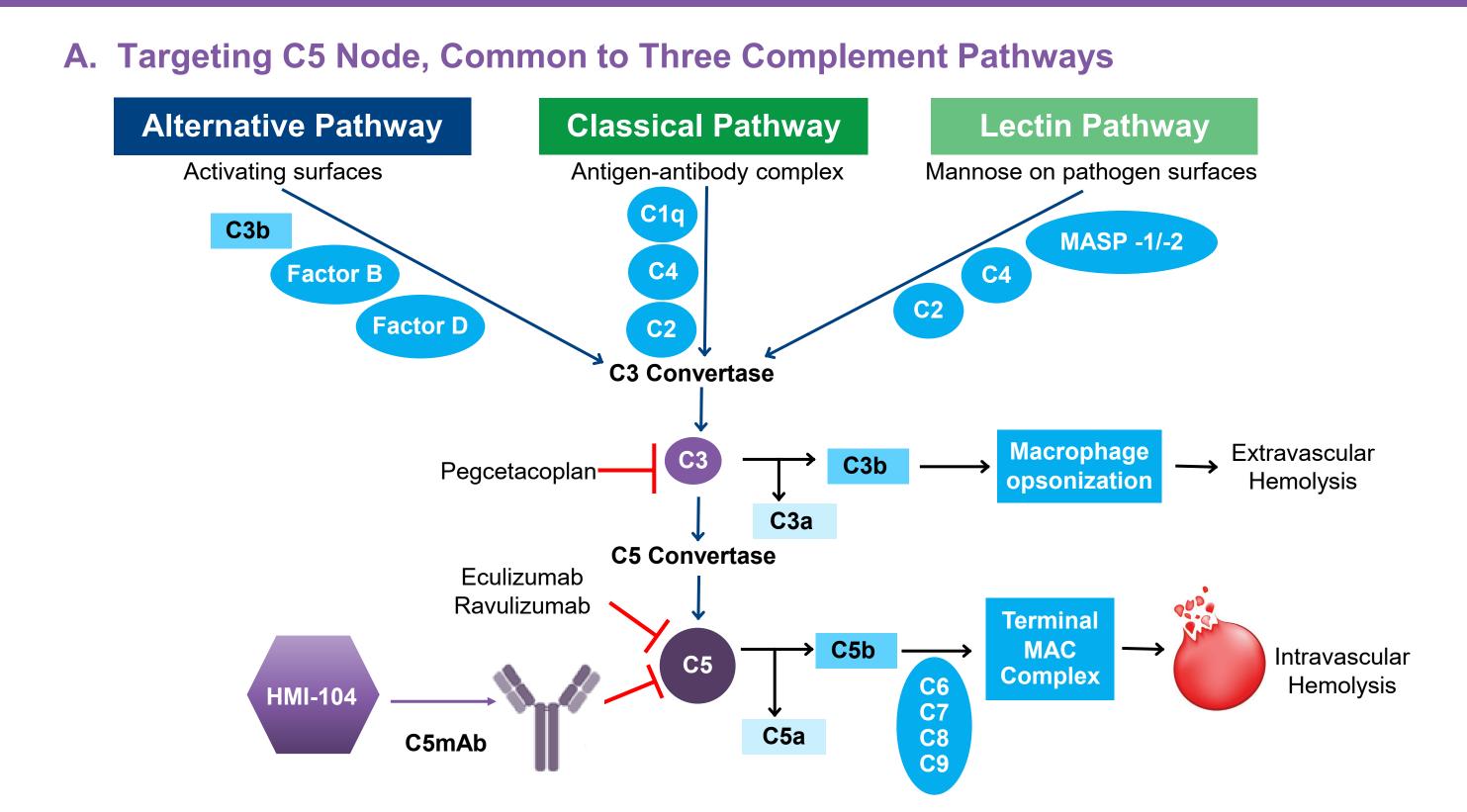
- HMI-104 successfully transduced human hepatocytes in vivo and expressed sustained serum C5mAb levels in the presence of human C5
- C5mAb expressed in the presence of human C5 was functionally active at all doses and timepoints tested, as determined by complete inhibition of ex vivo hemolysis using serum from HMI-104 treated FRG® KO mice
- Vgs and transcript levels in purified human hepatocytes were dose dependent
- ✓ Antibody levels achieved in FRG® KO mice were greater than or equal to those obtained at steady state in the NOD SCID mouse model

References: (1) Sharma, Y, et al. "Transducing the Liver as an Antibody Factory Using AAVHSCs." Molecular Therapy. Vol. 29. No. 4. 50 Hampshire St, Floor 5, Cambridge, MA 02139 USA: Cell Press, 2021. (2) FRG® KO Model details (https://www.yecuris.com/frg-ko-mice/). (3) Hill, A, et al. "Paroxysmal nocturnal haemoglobinuria." Nature reviews Disease primers 3.1 (2017): 1-14. (4) Gérard, S, et al. The Lancet 348.9027 (1996): 573-577. (5) Kelly, R. J., et al. Blood, The Journal of the American Society of Hematology 117.25 (2011): 6786-6792.

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HMI-104 is a Single Dose GTx-mAb that Expresses a Vectorized Monoclonal Antibody Against C5 for the Treatment of PNH

(work done at Yecuris)



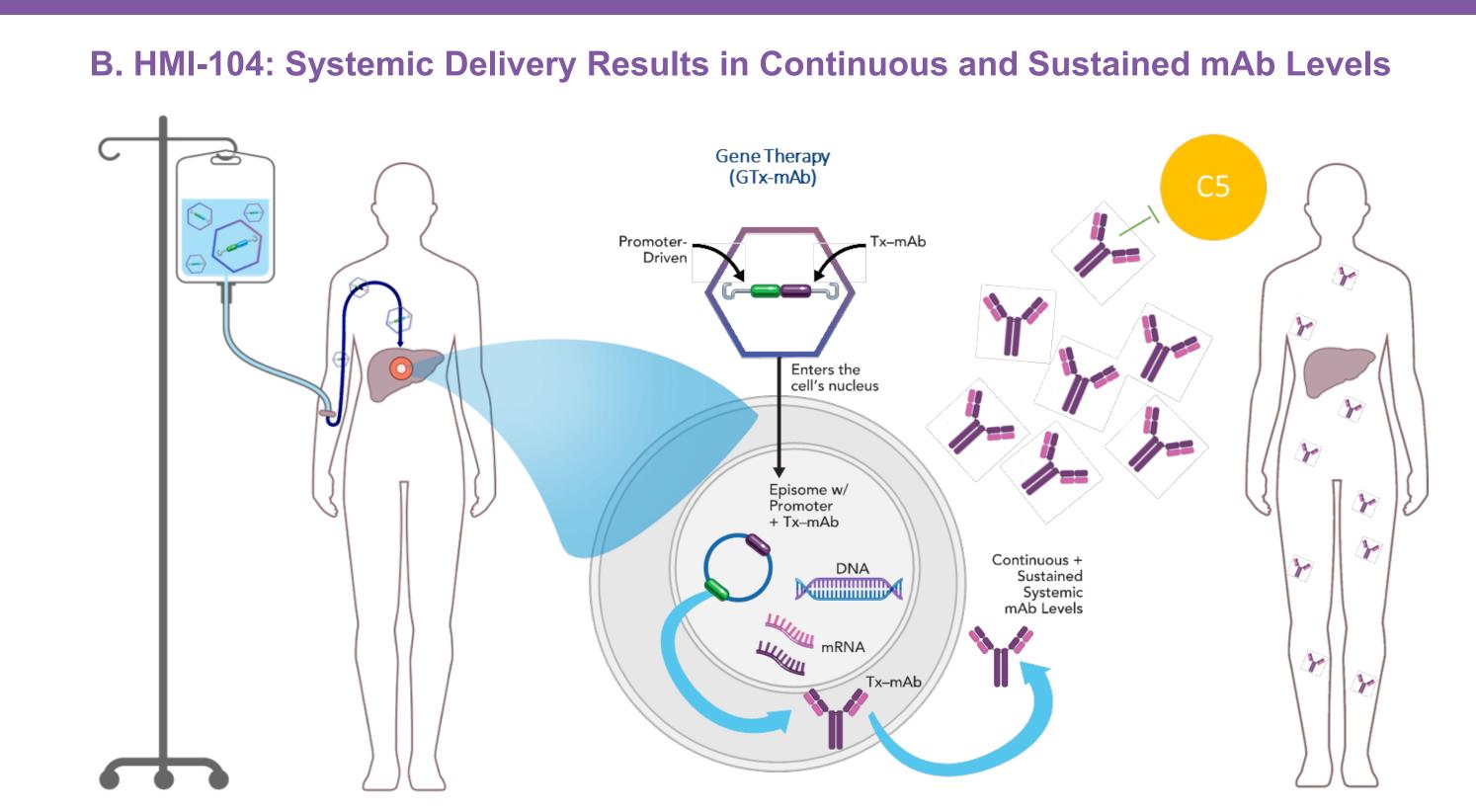
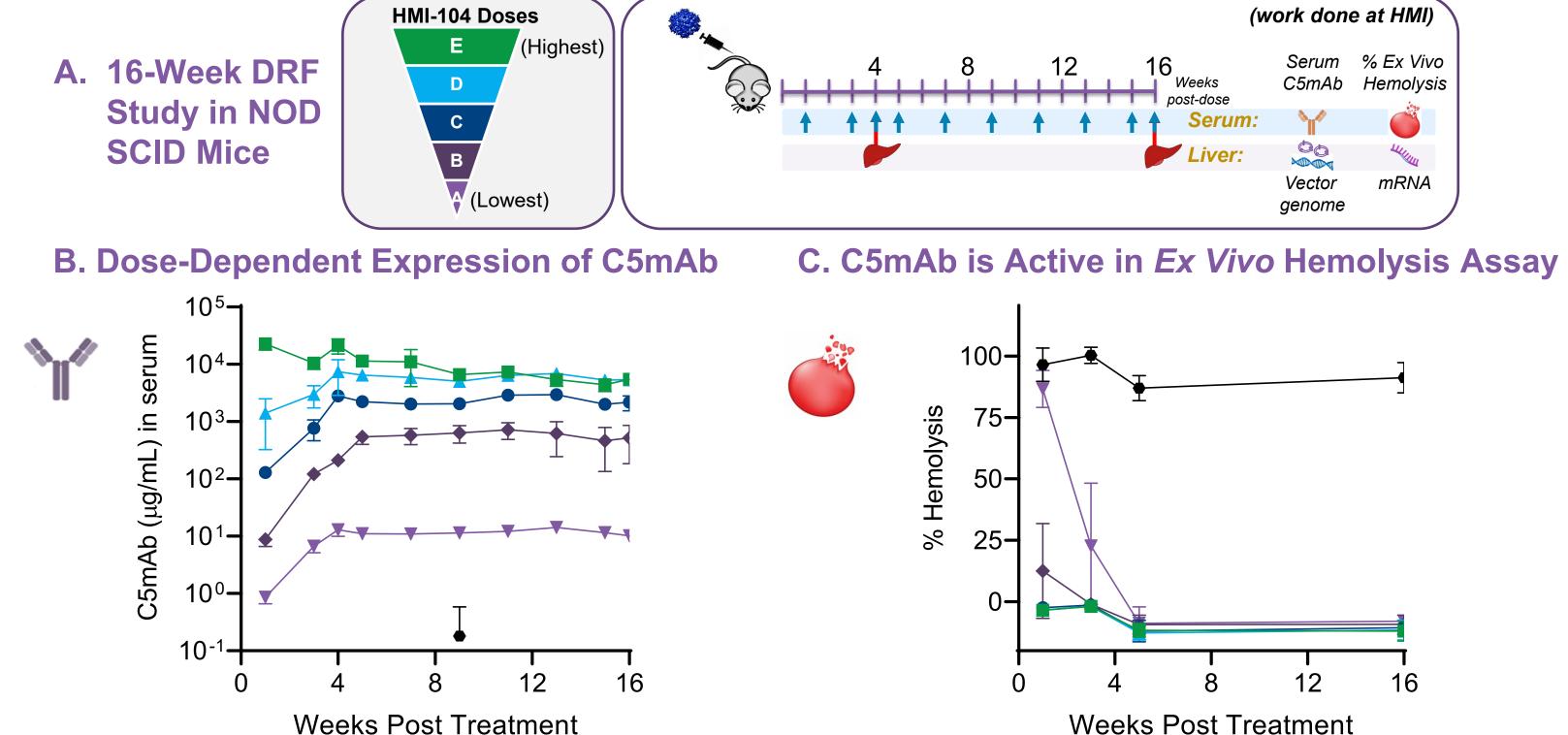


Figure 1: (A) Complement cascade highlighting approved C5 and C3 inhibitors and HMI-104 GTx-mAb approach; (B) HMI-104 is developed for one-time I.V. dosing, using the liver as an antibody factory to produce sustained expression of a C5mAb.

Treatment of NOD SCID with HMI-104 Results in Sustained, Dose-Dependent Expression of a Functional C5mAb, and Durable Liver VG and mRNA Levels



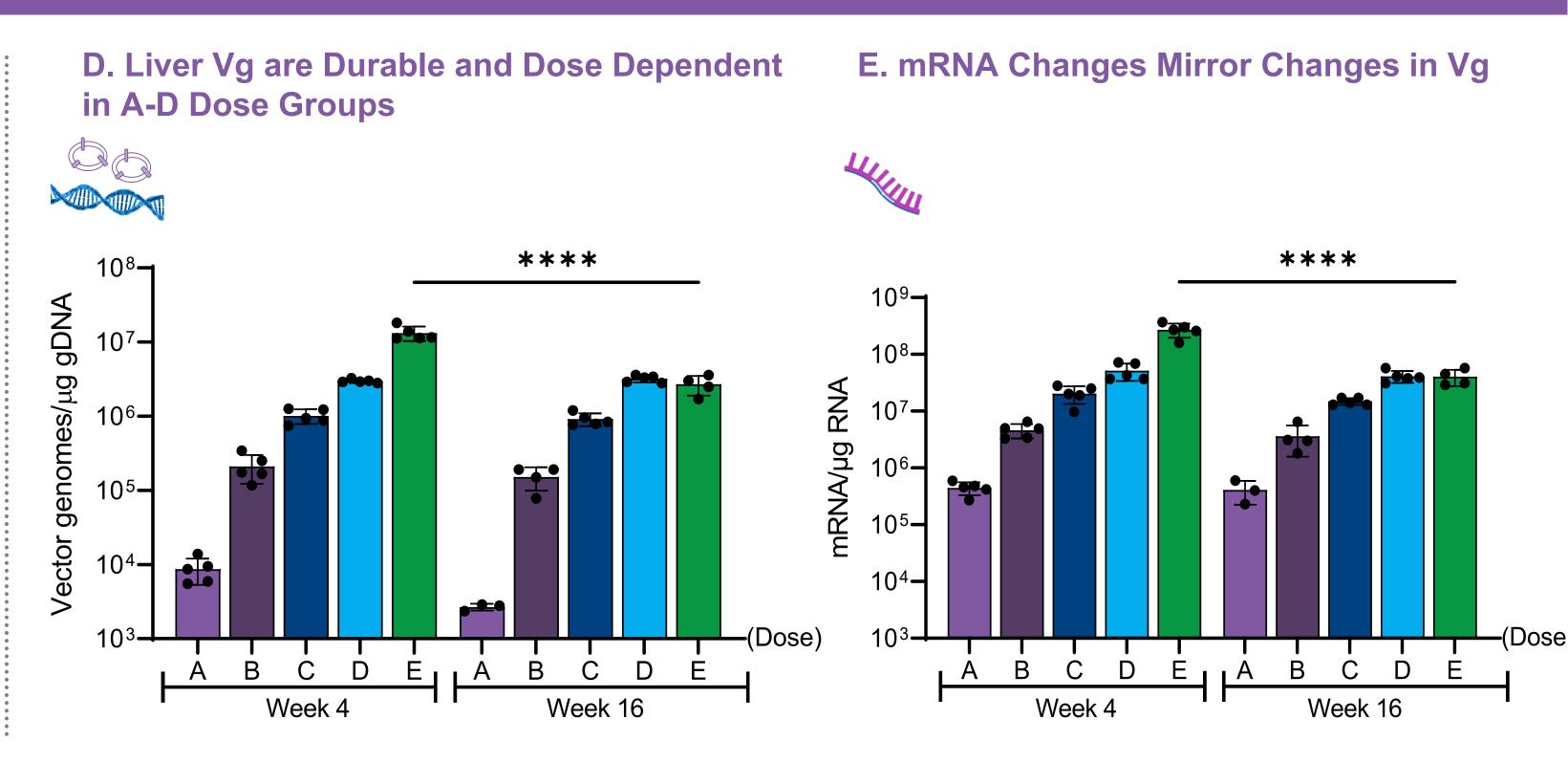
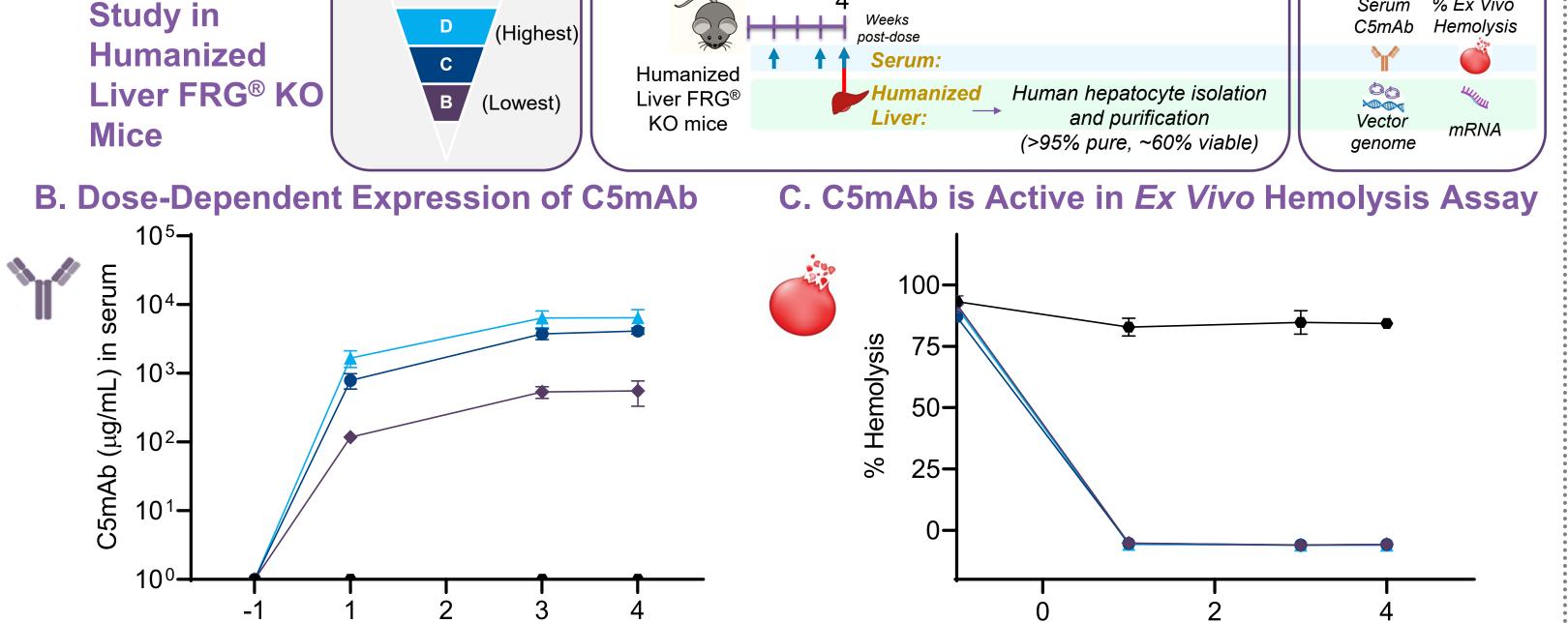


Figure 2: (A) 16-week study in NOD SCID male mice treated with HMI-104; (B) Time course of C5mAb levels in serum of NOD SCID mice following administration of HMI-104 at 5 doses (A-E) or vehicle control; (C) Inhibition of ex vivo hemolysis with serum from HMI-104-treated NOD SCID mice as a function of dose and time; (D) Dose-dependent changes in vgs and (E) mRNA in livers of HMI-104 treated mice. (Data represented as mean ± S.D.)

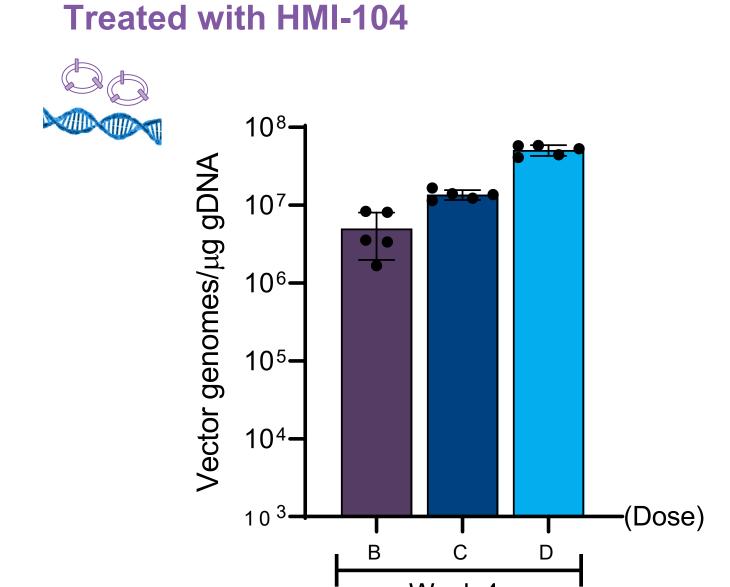
Treatment of Humanized Liver FRG® KO Mice with HMI-104 Results in Dose-Dependent Expression of a Functional C5mAb, with VG and mRNA in Human Hepatocytes

(work done at HMI)

Serum % Ex Vivo



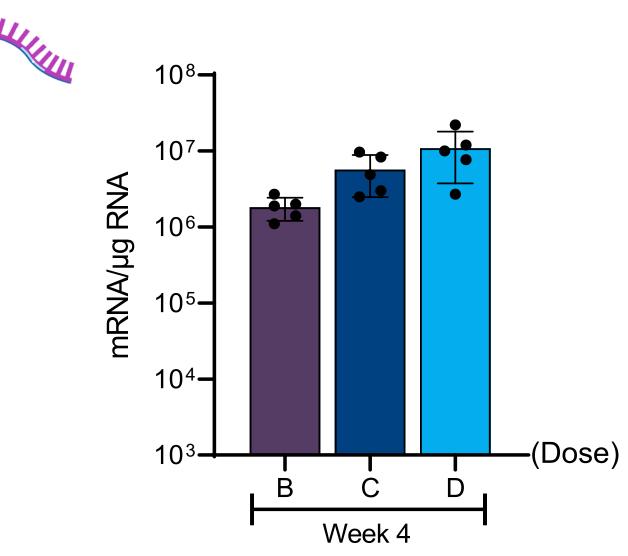
A. 4- Week DRF



D. Dose-Dependent Changes in Vg in Purified

Human Hepatocytes from FRG® KO Mice

E. Dose-Dependent Changes in mRNA in Purified Human Hepatocytes from FRG® **KO Mice Treated with HMI-104**



Weeks Post Treatment Figure 3: (A) 4-week study in humanized liver FRG® KO mice treated with HMI-104; (B) Time course of C5mAb in serum of FRG® KO mice following administration of HMI-104 at 3 Doses (B-D) or vehicle control; (C) Inhibition of ex vivo hemolysis with serum from HMI-104-treated FRG® KO mice as a function of dose and time (humanized liver FRG® KO mice express human and mouse C5, so assay adapted to inhibit murine C5; (D) Dose-dependent changes in vgs and (E) mRNA in human hepatocytes of HMI-104 Treated Mice. (Data represented as mean ± S.D.)

Conclusions

In conclusion, we present nonclinical data with HMI-104 demonstrating sustained expression of functional C5mAb levels in NOD SCID and humanized liver FRG® KO mice. These results support the development of HMI-104, which is currently in IND-enabling studies, for the treatment of PNH and complement-mediated disorders. Given the severity of PNH and the unmet need associated with available therapies, HMI-104 aims to provide, with a single I.V. treatment, sustained serum C5mAb sufficient to inhibit C5 complement-mediated lysis and reduce breakthrough and residual intravascular hemolysis associated with insufficient C5 antibody levels in PNH patients.